

From success to persistence: Identifying an evolutionary regime shift in the diverse Paleozoic aquatic arthropod group Eurypterida, driven by the Devonian biotic crisis

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Mass extinctions have altered the trajectory of evolution a number of times over the Phanerozoic. During these periods of biotic upheaval a different selective regime appears to operate, although it is still unclear whether consistent survivorship rules apply across different extinction events. We compare variations in diversity and disparity across the evolutionary history of a major Paleozoic arthropod group, the Eurypterida. Using these data, we explore the group's transition from a successful, dynamic clade to a stagnant persistent lineage, pinpointing the Devonian as the period during which this evolutionary regime shift occurred. The late Devonian biotic crisis is potentially unique among the "Big Five" mass extinctions in exhibiting a drop in speciation rates rather than an increase in extinction. Our study reveals eurypterids show depressed speciation rates throughout the Devonian but no abnormal peaks in extinction. Loss of morphospace occupation is random across all Paleozoic extinction events; however, differential origination during the Devonian results in a migration and subsequent stagnation of occupied morphospace. This shift appears linked to an ecological transition from euryhaline taxa to freshwater species with low morphological diversity alongside a decrease in endemism. These results demonstrate the importance of the Devonian biotic crisis in reshaping Paleozoic ecosystems.

KEY WORDS: Biotic crises, macroecology, macroevolution, mass extinction, morphological disparity, phylogeny.

Mass extinctions are known to be extraordinary events during which the normal rules of natural selection do not apply, and it has been suggested that a different selective regime acting upon cladelevel properties operates during these periods (Jablonski 2005). This suggests that taxonomic survivorship fits the paradigm of "nonconstructive selectivity" (Raup 1984) caused by nonrandom extinction but with survivorship determined by traits that are not otherwise strongly selected for during periods of background extinction. Of the "Big Five" Phanerozoic mass extinctions (Raup and Sepkoski 1982; Sepkoski 1986), the late Devonian is unusual in being characterized by a drop in speciation rates rather than any increase in extinction (Bambach et al. 2004), although two discrete extinctions, the Kellwasser and Hangenberg Events at the beginning and end of the Famennian (House 1985, 2002), have been posited. Causal factors for these events remain elusive (Chen et al. 2005), although marine transgression and widespread anoxia are frequently evoked (Bond and Wignall 2008; Carmichael et al. 2016). The cause of the anoxia is thought to be increased productivity due to the influx of nutrients from escalated continental weathering, resulting in eutrophication of the extensive shallow seas (Kaiser et al. 2016; Ma et al. 2016; Rakocinski et al. 2016). However, anoxia was prevalent before (Boyer et al. 2014) and after (Zaton et al. 2014) both the Kellwasser and Hangenberg Events, while some localities exhibit no evidence of anoxia during the events themselves (George et al. 2014), suggesting that anoxia alone could not be responsible for any mass extinction events. The rapid onset of glaciation during the shift out of greenhouse conditions, which prevailed through most of the Devonian (Slavik et al. 2016), has recently been suggested to be the driving factor for the late Devonian extinctions (McGhee et al. 2013) with cooling driven by the radiation of terrestrial plants (Myrow et al. 2014). Meanwhile, selenium depletion has been found associated with a number of extinction events throughout the Phanerozoic, including horizons at the end-Emsian and during the Famennian (Long et al. 2016). The pervasive nature of anoxia and repeated periods of selenium depletion throughout the Middle and Upper Devonian suggests a protracted period of environmental stress from the Emsian onwards with further perturbations such as rapid glaciation adding stress to an ecosystem already in crisis. Furthermore, biogeographic and phylogenetic studies of Laurentian species have suggested that the reduction in speciation is due to a decrease in provincialism caused by increases in interbasinal species invasions (Abe and Lieberman 2009; Stigall 2012). It has been proposed that these invasions were facilitated by transgressive marine pulses that enabled range expansion of ecological generalists, thereby eliminating vicariant speciation (May 1995; Abe and Lieberman 2009; Stigall 2012). Understanding the processes occurring during the late Devonian could, therefore, be important for modern conservation efforts in view of current fears of a human-driven sixth mass extinction (Barnosky et al. 2011) and faunal homogenization through biotic invasions and habitat loss (McKinney and Lockwood 1999).

Eurypterids are aquatic arthropods that form important components of Paleozoic ecosystems and are known to fill a variety of ecological roles (Anderson et al. 2014; McCoy et al. 2015). The group has a well-resolved phylogeny (Lamsdell 2013; Lamsdell et al. 2015), an evolutionary history ranging from the Middle Ordovician (467 mya) to the late Permian (254 mya), and comprises two constituent clades (Eurypterina and Stylonurina). Eurypterids, therefore, present an ideal case study for evaluating the impact of Paleozoic mass extinctions, particularly the late Devonian, which is known to have severely impacted the group (Lamsdell and Braddy 2010). Prior to the Devonian, eurypterids had successfully weathered one mass extinction event and appeared to be a thriving, successful group. Studying their decline, therefore, affords a rare opportunity to observe the ecological and evolutionary collapse of an otherwise globally dominant faunal component.

In this article, we analyze trends in taxonomic diversity and morphological disparity of the two eurypterid clades, Stylonurina and Eurypterina, along with rates of speciation and extinction across their recorded history. We explore the timing and impact of the Devonian biotic crisis, and compare it to the end-Ordovician and Kasimovian extinction events to examine how different forcing mechanisms influenced the trajectory of eurypterid evolution. In this way, we use past mass extinctions as natural experiments to better understand the processes occurring during our current biodiversity crisis.

Methods phylogenetic data

For the study, recent phylogenetic matrices for the Stylonurina (Lamsdell 2013) and Eurypterina (Lamsdell et al. 2015) were used. Both analyses were expanded to make species sampling more comprehensive and then combined into a single matrix, resulting in an analysis comprising 226 characters coded for 150 taxa. Of the 250 currently valid species of eurypterids (Dunlop et al. 2015), 145 were included in the phylogenetic analysis, of which 106 were Eurypterina and 39 were Stylonurina (see the data matrix; the data matrix is also available in the online MorphoBank database [O'Leary and Kaufman 2012] under the project code p2384, accessible at http://morphobank.org/permalink/?P2384, and through DRYAD at http://dx.doi.org/10.5061/dryad.24767). The 105 eurypterids not included in the phylogenetic analysis comprise taxa that are extremely poorly known and fragmentary, commonly consisting only of isolated carapaces or appendages. The majority of eurypterid higher taxa were represented in the analysis, with a few exceptions: Dorfopterus, which is known only from an isolated telson and may not represent a eurypterid (Tetlie 2007a; Lamsdell and Selden 2013); Eocarcinosoma, known from a single small carapace (Caster and Kjellesvig-Waering 1964); Marsupipterus, also known from a single telson but which has been considered to either show similarities to basal Eurypterina (Braddy et al. 1995; Lamsdell 2011) or be a phyllocarid crustacean (Rolfe 1963); Necrogammarus, previously considered to be a crustacean or myriapod but since shown to be a pterygotid eurypterid likely synonymous to either Pterygotus or Erettopterus (Selden 1986); and Unionopterus, which is some form of adelophthalmoid, although the original description is somewhat lacking and the holotype and only known specimen is currently lost (Tetlie and Van Roy 2006).

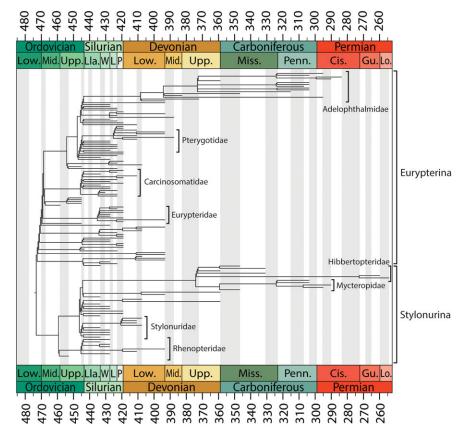


Figure 1. Phylogenetic tree of Eurypterida used in the diversity and disparity analyses. Tip labels excluded for clarity (but see Fig. S1), although clades mentioned in the text are indicated. Although basic range extension was used in all analyses, a minimum branch length of 1 million years was used to aid in interpretation of tree topology in this figure.

Individual species were coded to remove the potential for the inclusion of paraphyletic or polyphyletic groups. The application of paraphyletic or polyphyletic groups in biodiversity studies can have drastic effects on our understanding of mass extinction events. Erroneous patterns of pseudoextinction can be retrieved when monophyly is not recognized, whereas incorrect assumptions of monophyly can dampen the observed loss of biodiversity when studies are conducted at higher taxonomic levels (Patterson and Smith 1989). The data matrix was subjected to cladistic analysis using TNT (Tree analysis using New Technology: Goloboff et al. 2008), employing random addition sequences followed by tree bisection-reconnection (TBR) branch swapping with 100,000 repetitions with all characters unordered and of equal weight (for a recent review of the issues surrounding differential character weighting see Congreve and Lamsdell 2016). The strict consensus tree was constructed (Figs. 1 and S1) and used as the framework for all subsequent diversity and disparity analyses.

DIVERSITY

The phylogenetic tree was plotted against geological time, based on an existing eurypterid database (Lamsdell and Braddy 2010), updated for recent discoveries. Taxon occurrence was extended according to implied ghost ranges (Norell 1992) and parsimony reconstructed ancestors included as inferred from the tree topology (Brusatte et al. 2011), thereby correcting for potential stratigraphic biases (see discussion of phylogenetic diversity estimates [PDEs] next). Where reconstructed character states conflicted between different optimization models (accelerated or decelerated transformation), they were considered equivocal and coded as unknown. These inferred ancestors were assigned ranges based on the basic time-scaling methods of Norell (1992) and Smith (1994). Although this method can result in zero-duration branch lengths (Bell and Lloyd 2015), this is a nonissue for the current study as analyses requiring some estimate of branch length, such as rates of morphological evolution, were not performed. Basic time-scaling methods are acceptable protocols for identifying times of divergence and assessing relative variation in diversity (Bapst 2014), in which they outperform minimum branch length scaling methods (e.g., Brusatte et al. 2008). This resulted in the earliest occurrences being capped at the Darriwilian, in accordance with the occurrence of the earliest known eurypterid fossils (Lamsdell et al. 2015). Although pre-Darriwilian eurypterids most likely exist, given that the earliest recorded eurypterids are not the most phylogenetically basal, this measure affects a most conservative estimate of eurypterid diversity throughout the Paleozoic. Two measures of diversity were calculated, a taxic diversity estimate (TDE; see Levinton 1988) derived from the raw species counts and a PDE (Norell 1993) incorporating ghost range extensions and inferred ancestors based on the phylogenetic tree, for stagelevel time intervals (Table S1). A number of processes-such as uneven geographic sampling, unequal outcrop availability, and differential preservation potentials in different depositional environments (Newell 1959; Simpson 1960; Raup 1972)-are known to cause inherent bias in the fossil record, and so corrections must be applied to the raw diversity counts to attain a numerical estimate of the relative amount of diversity in each time interval (Alroy 2010a). When only bulk diversity counts are available, methods such as shareholder quorum sampling (SQS) can be used to dampen the impact of these biases (Alroy 2010b); however, when a phylogenetic framework is available, this can be used to generate a PDE to provide an estimate of minimum relative diversity (Norrell 1992, 1993; Smith 1994). Although a fundamentally different approach than SQS, PDEs also reduce the impact of sampling biases and have been shown to outperform TDEs using both simulated (Lane et al. 2005) and actual (Butler et al. 2009) datasets. PDEs are known to exhibit some artifacts due to the Signor-Lipps effect, whereby the terminal portion of a taxon's range is truncated through increasing rarity of individuals decreasing the probability of sampling (Signor and Lipps 1982; Wagner 2000), thereby dampening the impact of abrupt mass extinction events (Lane et al. 2005). However, SOS is equally impacted by the Signor-Lipps effect (Alroy 2014) and alternative methods such as use of the three-timer (3T) log ratio (Alroy 2010c) are noisy when turnover rates are high (Alroy 2014), as would be expected with species-level data and stagelevel time bins. Furthermore, it has been noted that the record of mass extinctions is biased toward widespread, dominant taxa, and that apparent extinctions in the fossil record often mark critical drops in abundance (Hull et al. 2015), a phenomenon that is also observable when looking at the record of modern threatened and endangered species (Plotnick et al. 2016). Failure to sample the termination of a lineage may then be widespread and therefore not impart as much of a directional bias into diversity studies as feared, while it can also be argued that terminally dwindling populations are already functionally extinct from an ecosystem perspective. It has also been shown that gradual extinctions can appear abrupt (Wang et al. 2012), sometimes mediated by clustering of last observed occurrences at sequence stratigraphic horizons mediated by sedimentary hiatuses and changes in water depth (Holland and Patzkowsky 2015). The Signor–Lipps effect, therefore, appears to be less ubiquitous, and thus less directionally biasing, than initially conceived. Similar biases caused by Lazarus taxa-taxa that dwindle in abundance resulting in apparent extinction only to recover and reappear in the fossil record (Jablonski 1986)-that

the 3T sampling method explicitly seeks to remove are also accounted for in PDEs as any gaps in the ranges of the operational taxonomic units (OTUs—in this case, species) are filled as part of ghost range extension.

Taxic diversity, including that derived through PDEs, can also be biased by the Lagerstätten effect, whereby sites of exceptional preservation inflate relative abundance through increased sampling (Butler et al. 2009). These artifacts are, in part, thought to be caused by taxonomic oversplitting of exceptionally preserved material; however, much work has been done in recent years to reduce this phenomenon among eurypterids (e.g., Braddy 2000; Tetlie 2006; Lamsdell 2011). A number of taxa that are likely synonymous but have yet to be officially mergedfor example, Parahughmilleria hefteri and Parahughmilleria major (Størmer 1973), considered to be ontogenetic variants of one another; Moselopterus ancylotelson and Moselopterus elongatus (Størmer 1974), M. elongatus probably representing an undeformed specimen of M. ancylotelson; and Eurypterus remipes and Eurypterus lacustris, considered chronospecies by Tetlie et al. (2007) and therefore synonyms according to any species definition criterion-were treated as synonyms for all analyses. Taxonomic oversplitting is therefore not likely to bias the current study, and so the impact of the Lagerstätten effect will be dampened. Comparing the eurypterid diversity estimates with a recent compendium of fossil Lagerstätten (Retallack 2011) shows that, aside from large peaks in the Cambrian (a period from which no eurypterids are known), overall Lagerstätten frequency has remained relatively constant throughout the Phanerozoic. When narrowing the comparison to only localities preserving cuticle and nonbiomineralized soft tissue, the post-Cambrian Paleozoic record is again shown to be relatively constant, with the exception of notable peaks in the late Silurian and late Devonian. The Devonian peak does not manifest in the eurypterid diversity estimates; however, the late Silurian peak is apparent in the TDE, but not the PDE as it is subsumed into a generally high Silurian diversity (Fig. 2). Diversity estimates of late Silurian eurypterids may therefore be magnified in the TDE; however, the lack of a corresponding peak in diversity during the late Devonian uptick in Lagerstätten occurrence suggests the decrease in eurypterid abundance during the Devonian is real. It is worth noting also that eurypterids are explicitly used as indicators of Lagerstätten-type preservation (Retallack 2011) and so the late Silurian peak in exceptional preservation occurrence is largely driven by an increase in the number of eurypterid localities; as such, it is impossible to separate whether this increase is due to an increase in suitable preservational environments or simply because eurypterids are more abundant at this time. Furthermore, eurypterid occurrences are not limited solely to sites of exceptional preservation, and eurypterids are frequently preserved throughout their geologic history in a variety of lithologies, including sandstones, where cuticle is ephemeral or entirely

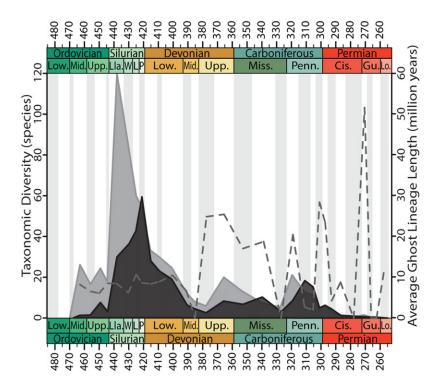


Figure 2. Species-level diversity curves for Eurypterida (derived from the data in Table S1). Taxonomic diversity estimate (TDE) is shown in black, phylogenetic diversity estimate (PDE) in gray. Average ghost lineage duration is indicated by the dashed line.

absent (e.g., Cope 1886; Kjellesvig-Waering 1950, 1961; Leutze 1961; Lamsdell et al. 2009; Lamsdell et al. 2013a,b; Budil et al. 2014). As such, the increase in eurypterid abundance during the Silurian is considered to be a real signal, although the magnitude of this increase may be somewhat exaggerated. To test this, average ghost lineage duration (Cavin and Forey 2007) was calculated for each time bin to explore which peaks in diversity result in a reduction in average ghost range duration (representing a genuine biological radiation) and which result in no reduction in average ghost range duration and therefore are a result of the Lagerstätten effect. Cavin and Forey, however, assumed a "budding" mode of speciation, arbitrarily assigning the range of the inferred ancestral taxon to the basal-most member of a clade-something that may prove impossible if the clade does not have a pectinate topology. To circumvent these issues, we simply accounted for the inferred ancestral taxon as a separate ghost range at the earliest occurrence of the clade itself.

A final concern, that PDEs assume a "true" cladogram for the study group exists (Lane et al. 2005), is refutable based on the fact that no cladogram is ever considered to represent the "true" tree of organism relationships but is instead a testable hypothesis based on the available data and can be further refined in the future. In this manner, it is no different than studies that use SQS and 3T metrics, as these are also hypotheses about taxon distributions given the available data. If multiple hypotheses (cladograms) existed for the relationships of the study group then the analyses could be performed on all the available cladograms and compared, or cladograms could be evaluated for inclusion prior to the analysis. In this case, however, only a single dataset exits, although analysis of this dataset does result in multiple cladograms. As noted above, these cladograms are summarized through strict consensus and this subsequent tree was used as the framework for this study so as not to bias the results toward any of the equally supported most probable trees. Because conflicts are summarized through polytomies, this results in the loss of an inferred ancestor and so represents the most conservative reconstruction of unsampled diversity and disparity. Both TDE and PDE measures show congruent patterns of diversity shifts across the Paleozoic (Fig. 2), and so the PDE was used for the diversity/disparity comparisons. Using the PDE, estimates of diversity are directly comparable to the disparity data, which also incorporates ghost range extensions and inferred ancestral taxa. The results and interpretations are not altered by using the TDE, nor when excluding inferred ancestral taxa from the disparity analyses.

RATES OF SPECIATION AND EXTINCTION

Rates of speciation and extinction were calculated for each clade at the stage level from the PDE range data. Numerous speciation and extinction metrics have been proposed for rate calculation of modern and fossil taxa, most commonly based on the deterministic exponential model of taxon growth (Sanderson and Donoghue 1996; Nee 2004, 2006). Rates calculated from paleontological data are commonly derived from datasets without any phylogenetic structure that generate per-capita rates normalized for standing diversity (Alroy 2014). The most widespread of these methods for rate calculation is the generalized boundary-crosser (BC) method (Foote 2000a,b) that incorporates counts of taxa either crossing both the bottom and top boundaries of an interval or crossing one boundary but not the other. However, as with raw counts of taxic diversity, the BC method can be influenced by the occurrence of Lazarus taxa and other sampling biases, and alternative methods such as the 3T (Alroy 2010c) and gap filler (GF; Alroy 2014) equations have been proposed to ameliorate these effects. Despite this, simulations have shown that much of the bias in BC analyses stems from the use of higher taxa, with taxonomic practices of overlumping of long-ranging, rare genera biasing the results (Alroy 2014)-an issue largely circumvented by the use of species-level data. The issue of Lazarus taxa is again removed through the utilization of phylogenetically corrected ranges. Deterministic rates have also been used in paleobiological analyses where phylogenetic frameworks are available (Hulbert 1993; Rode and Lieberman 2005; Abe and Lieberman 2009; Stigall 2010). These rates are derived from the standard equation for exponential growth as first used by Maurer (1989)and therefore are not modifications to Foote's (2000a, b) BC equation as claimed by Rode and Lieberman (2005) and Abe and Lieberman (2009). These methods use the phylogenetic tree to correct for sampling biases and remove the effect of Lazarus taxa (Smith 1994), and are more suitable for species-level data, which are more likely to result in short-ranged taxa with a greater preponderance of singletons.

Comparison of calculated rates allows a broader basis for interpreting the generality of observed biodiversity patterns (Stigall 2010). Per-capita rates for speciation (\hat{p}) and extinction (\hat{q}) were calculated for each time bin following the BC method of Foote (2000a,b):

$$\hat{p} = -\ln \left(N_{bt} / N_t \right) / \Delta t,$$
$$\hat{q} = -\ln \left(N_{bt} / N_b \right) / \Delta t,$$

where N_{bt} is the number of taxa that cross both the lower and upper boundary of the time bin, N_b is the number of taxa that cross the lower time bin boundary from the previous time bin, N_t is the number of taxa that cross the upper time bin boundary and persist into the subsequent time bin, and Δt is the length of duration (in million years) of the time bin. Deterministic rates of speciation rate (*S*) and extinction rate (*E*) were calculated using the equations of Maurer (1989) as set out by Hulbert (1993):

$$S = (\ln (N_0 + o_0) - \ln N_0) / \Delta t,$$

$$E = (\ln (N_0 + o_0) - \ln N_1) / \Delta t,$$

where N_0 is the number of taxa that cross the lower time bin boundary from the previous time bin, N_1 is the number of taxa that cross the upper time bin boundary and persist into the subsequent time bin, o_0 is the number of taxa that originate within the time bin, and Δt is again the length of duration (in million years) of the time bin. Therefore, N_0 is equivalent to N_b , N_1 is equivalent to N_t , and N_0 combined with o_0 is equivalent to Foote's (2000a,b) N_{tot} , representing the total diversity of a time bin. Both methods suffer from edge effects caused by the sudden origination or extinction of a large amount of taxa in the first and last time bins, respectively; while an edge effect is unlikely in the last time bin (the Wuchiapingian) as eurypterid diversity has already drastically winnowed down to only a handful of species, an edge effect is likely in the first time bin (the Darriwilian), and this should be considered when reviewing the rates retrieved for the Darriwilian.

DISPARITY

Analyses of disparity (morphological diversity) have become increasingly common in the paleobiological literature over the last couple of decades, in particular those utilizing discrete character matrices (Foote 1994, 1999; Wills et al. 1994; Wills 1998; Lofgren et al. 2003; Wesley-Hunt 2005; Brusatte et al. 2008; Young et al. 2010; Thorne et al. 2011; Bapst et al. 2012; Hughes et al. 2013; Ruta et al. 2013; Hopkins and Smith 2015; Lamsdell 2016; Oyston et al. 2016). These studies have employed a number of different distance metrics to generate Euclidean pairwise distances between all taxa (see Lloyd 2016); however, the two most commonly used are generalized Euclidean distance (GED: Wills et al. 1994) and Gower's coefficient (GC: Gower 1971). The two metrics differ primarily in their handling of missing data, with GED inserting a weighted mean fractional univariate distance based on those distances that are calculable, while GC simply rescales calculable distances based on the amount of information available. Data are handled during this process in a phenetic manner, with no assumptions of hierarchy or homoplasy, and as such it is not circular to compare results from disparity analyses to the phylogenetic hierarchy retrieved from the same character matrix. Analyses of disparity formulated from discrete character matrices have been shown to converge on a common signal with other forms of morphometric data (Foth et al. 2012; Hetherington et al. 2015) and so results are comparable with those attained from, for example, landmark data.

Eurypterid disparity measures were calculated from the cladistic character data through GED. Along with the observed species, reconstructed ancestors (Brusatte et al. 2011) were included as necessitated by tree topology. Ancestors were coded into the phylogenetic matrix with their characters reconstructed using parsimony-based ancestral state reconstruction in Mesquite (Maddison and Maddison 2015) and outgroup non-eurypterid taxa

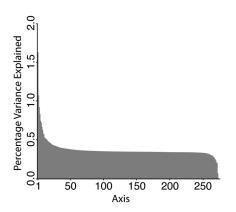


Figure 3. Percentage variance encompassed by each axis of the principal coordinates analysis (PCO).

(Weinbergina and the chasmataspidids Chasmataspis, Loganamaraspis, Octoberaspis, and Diploaspis) were removed from the matrix prior to analysis. Autapomorphies (character states unique to single taxa) were included as, unlike in parsimony analyses where they contribute no information to the inference of relationships, unique character states can be important for determining the relative distance between taxa in morphospace. However, with the inclusion of inferred ancestors and the fact that autapomorphies were frequently coded as a single state within a multistate character, the only continuous characters in the analysis were those rendered invariable after the removal of outgroup taxa, and so these characters were also excluded from the analysis.

In the resulting matrix, 41% of the character codings were missing or inapplicable, although these were not evenly distributed throughout the matrix, resulting in some taxa having significantly fewer characters coded than others. This resulted in GED being used over GC, as GC will result in incalculable distances if there are few comparable characters for a specific distance, whereas GED ensures that all taxa are included in the analysis as long as there is at least one coded character in common between two taxa. Pairwise (taxon to taxon) distances were calculated with negative eigenvalue results for some axes removed through the addition of a constant, calculated computationally following Cailliez (1983), and subjected to ordination through principal coordinates analysis (PCO) using scripts in R (Hughes et al. 2013). One characteristic of analyses of discrete character data is that the variance tends to get spread relatively evenly over each of the axes (Lloyd 2016); as such, it can be more informative to look for discontinuities among the distribution of encompassed variance per axis. Three discontinuities are evident on the distribution of variance generated from the eurypterid data, one after the second axis, another after the fourth, and the last after the tenth (Fig. 3). However, these axes only represent 2.7, 4.4, and 8.3% of the total variance, respectively. As such, phylomorphospace plots were produced using the first two axes in Scatterplot3d (Liggs and

Maechler 2003), as these two axes are representative of the overall changes in spatial distribution that the morphospace exhibits in each time bin, however, all statistical tests were performed using data from all axes.

Disparity was quantified through the sum of range (SOR), which describes the total range of morphospace occupation represented by the volume of a hypercube that encompasses all of the morphotypes (Foote 1991) thereby showing the size of dissimilarity within a morphospace, and the sum of variance (SOV), which is the sum of variances along the morphospace axes (Van Valen 1974) and acts as a measure of the distances between the points (taxa) in morphospace. SOR and SOV were calculated for each time bin using all axes and presented as the mean value of 1000 bootstrap replicates; SOR and SOV values were considered distinct from one another if there was no overlap of their standard error (SE). Changes in morphospace occupation between time bins were quantified using the percentage difference in SOR, SOV, and position of the centroid (POC)-the median value along each morphospace axis (Dommergues et al. 1996)-resulting in percentage difference in SOR (P_{SOR}) , percentage difference in SOV (P_{SOV}), and percentage difference in POC (P_{DPOC}) values for each time bin pair. These statistics can be used to quantify whether changes in morphospace are random (nonselective) or marginal or lateral (selective; Korn et al. 2013; see Table S2). As originations were being examined as well as extinctions, the presentation of the resulting figures was modified from that in Korn et al., with 100% (no change) taken as a baseline and variations taken as a positive or negative percentage from that (therefore a value of 116% in Korn et al. would be +16% herein, while a value of 78% would be -22%), thereby permitting easier comparisons between extinctions and radiations. Although values for P_{SOR} and $P_{\rm SOV}$ are presented as either positive (representing an increase in SOR or SOV) or negative (representing a decrease), P_{DPOC} simply indicates any deviation across the sum of all axes, and therefore is directionless, and as such P_{DPOC} is shown as a positive value.

 P_{SOR} , P_{SOV} , and P_{DPOC} were calculated across the Katian/Hirnantian and Hirnantian/Llandovery stage boundaries, for the Kasimovian/Gzhelian stage boundary, and for each stage boundary from the Lochkovian to the Tournasian, encompassing both the onset and breakdown of late Ordovician glaciation (Sheehan 2001; Congreve 2013), the Kasimovian rainforest collapse (Sahney et al. 2010a), and the entire duration of the Devonian. Changes in morphospace occupation caused through extinction were calculated by comparing the occupied morphospace of a time bin with that of only surviving taxa in the succeeding time bin, whereas morphospace changes through origination were calculated by comparing the occupied morphospace of a time bin with that of taxa originating in the succeeding time bin. In this manner, we can quantify whether both origination and extinction

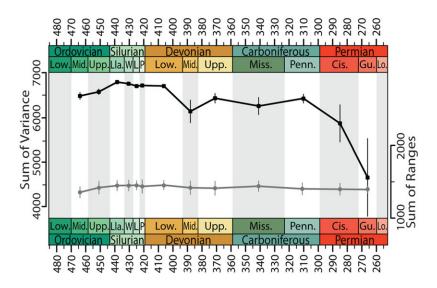


Figure 4. Disparity metrics for Eurypterida from the Euclidean distance matrix derived from the character matrix. Sum of variances (SOV) denoted by squares with black lines, sum of ranges (SOR) denoted by circles with gray lines. Error bars are 95% confidence intervals. Values are rarified to correct for nonstandard sample size, and epoch-level time bins were used.

are random or directional in regard to morphospace occupation, and whether any changes in morphospace occupation are driven by extinction, origination, or both. Multivariate statistical tests (PERMANOVA - permutational multivariate analysis of variance using the Euclidean distance measure) were performed to ascertain the statistical significance of overlap and separation of morphospace across all axes between time bins at the epoch and period level. Significance was estimated by permutation across groups with 10,000 replicates. Statistical analysis was performed both with and without Bonferroni correction. Bonferroni correction provides little statistical power and is probably too conservative (Sokal and Rohlf 1995; Moran 2003; Nakagawa 2004; Garamszegi 2006); instead, effect size is reported as a more accurate measure of robustness (Cohen 1988). Pairwise comparisons with Bonferroni correction generally exhibit less significance than those without, but still display the same general trend (Tables S3-S6).

Results and Discussion diversity, disparity, and shifts in morphospace occupation through time

Diversity and disparity are decoupled throughout the evolutionary history of Eurypterida, with disparity remaining relatively constant (Figs. 4 and S2) irrespective of increases and decreases in diversity (Fig. 2). Similar patterns have been shown in graptolites (Bapst et al. 2012) and vertebrates (Ruta et al. 2013), thus emphasizing the importance of considering both diversity and disparity in studies of clade history. Eurypterids exhibited low levels of diversity in the Ordovician (485.4–443.8 mya) before under-

Barbon StrengthEurypterids exhibited low levelsdwOrdovician (485.4–443.8 mya) before under-miEVOLUTION 2016

going a massive increase, reaching their peak diversity in the Silurian (443.8-419.2 mya), with decreases in average ghost range length during the Upper Ordovician and early Silurian showing that these radiations are at least in part real. Eurypterids do not appear to have been overtly impacted by the end-Ordovician mass extinction although rates of extinction underwent an increase in the Hirnantian (445.2-443.8 mya) that may correlate to the rapid onset and subsequent breakdown of extreme glaciation (Sheehan 2001; Congreve 2013). The clade exhibited a marked drop of diversity into the Devonian, driven in part by peaks of extinction in the Pridoli (423-419.2 mya) and Pragian (410.8-407.6 mya), with a subsequent steady decrease from the Emsian (407.6–393.3 mya) through to the Frasnian (382.7–372.2 mya). By the onset of the Carboniferous, only the Adelopthalmidae of the Eurypterina and the Hibbertopteridae and Mycteropidae of the Stylonurina persist, with previously dominant clades having declined to extinction by the Middle Devonian (Carcinosomatidae are last known from the Lochkovian, Stylonuridae from the Pragian, Rhenopteridae and Eurypteridae from the Emsian, and the Pterygotidae from the Eifelian). The SOV also dropped during the Middle Devonian (393.3-382.7 mya), although no corresponding drop is seen in the SOR; diversity and SOV subsequently remained at relatively low levels for the remainder of the Paleozoic before dwindling further during the Permian (298.9-252.2 mya), until the eventual extinction of the group in the Wuchiapingian (259.8-254.2 mya). Such a pattern is consistent with that of survival without recovery (Jablonski 2001), making post-Devonian Eurypterida a "dead clade walking" (Jablonski 2002), albeit one that dwindles to a reduced ecological niche rather than suffering imminent extinction.

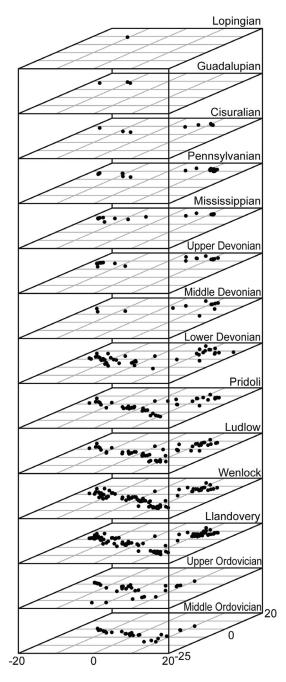


Figure 5. Morphospace occupation by Eurypterida throughout the Paleozoic. Plots are based on the first two axes from the principal coordinates analysis (PCO).

Morphospace occupation is shown to remain mutable throughout the majority of the Paleozoic (Fig. 5). By the Carboniferous, however, morphospace occupation had been significantly reduced, and stayed restricted for the remainder of the Paleozoic. These visual comparisons are confirmed statistically: PERMANOVA tests show that earlier Paleozoic morphospace occupation is significantly distinct from that in the late Paleozoic (Tables 1 and S7–S9), while the Carboniferous occupied morphospace is not significantly distinct from that in the Permian. Morphospace occupation is seen to shift during the Devonian, with the position of species in morphospace shifting to one of the lateral extremes during the Middle Devonian from a more central position during the Ordovician–Silurian. This correlates with the decrease in morphospace mutability, with the morphospace occupation of each epoch from the Middle Devonian onwards being statistically indistinguishable from one another (Tables S7 and S9). This shift is not reflected in the SOR (Figs. 4 and S2) as the overall range of morphospace occupied remains relatively unchanged, with two groups located in disparate regions of morphospace (Fig. 5); however, the amount of variance has decreased as the average distance between species decreases, with the two groups being tightly clustered; this is reflected in the decrease in SOV, which occurs during the Middle Devonian (Figs. 4 and S2).

Quantifying the change in morphospace occupation between stages due to extinction (Korn et al. 2013), however, reveals that extinction was random throughout the Devonian (Tables 2 and S10), with P_{SOR} remaining well below the >10 values that would indicate a nonrandom extinction, while P_{SOV} remains below the >13 value threshold and P_{DPOC} is fixed at around 1–2, well within random levels. Quantification of extinction during the twin pulses of the end-Ordovician mass extinction (Sheehan 2001; Congreve 2013), the Kellwasser and Hangenberg events (House 1985, 2002), and the Kasimovian rainforest collapse (Sahney et al. 2010a) reveals that random reduction of morphospace occupation is a consistent response among eurypterids during extinction events. The subsequent lateral shifts in morphospace occupation during the Devonian are therefore due to differential patterns of origination rather than extinction; origination became increasingly skewed throughout the Early Devonian, with P_{SOV} and P_{DPOC} values dropping at each subsequent stage boundary (Table 2 and S10). Negative values for P_{SOV} during origination indicate that the originating species are more clustered in morphospace than the species in the previous time bin, indicating that only a subset of morphospace was being repopulated by new species after the random extinction events. As the threshold values required for a meaningful shift in morphospace drop as the magnitude of change increases (Korn et al. 2013), it is significant that the P_{SOR} , P_{SOV} , and P_{DPOC} values for origination during the Devonian stages are consistently higher than those for extinction despite the magnitude of change due to origination being less than half of that due to extinction. Therefore, selective patterns of speciation, rather than extinction, were responsible for changes in morphospace occupation across the Devonian.

SHIFT IN EVOLUTIONARY REGIME AND THE DEVONIAN BIOTIC CRISIS

Eurypterids underwent a major shift in evolutionary regime during the Devonian, corresponding to the migration and subsequent

Table 1. PERMANOVA test results of Eurypterida ($F_{(5,321)} = 1.298$, $\eta^2 = 0.016$, P = 0.0001) from 100,000 permutations for statistical differences between taxa for each of the six period-level time bins based on PCO analyses.

	Ordovician	Silurian	Devonian	Carboniferous	Permian
Ordovician	_	1.1810	1.4900	1.7860	1.3390
Silurian	0.0009	—	1.2200	1.6610	1.2100
Devonian	0.0001	0.0004	_	0.9995	0.9998
Carboniferous	0.0001	0.0001	0.4798	—	0.7111
Permian	0.0001	0.0001	0.4244	0.9987	_

Nonsignificant *P* values (indicating no significant difference between time bins) shown in bold. Values in regular font are for the *P* values, those in italics are the raw *F* values.

Table 2. Quantification of changes in morphospace across stage boundaries (Korn et al. 2013). Entries in bold represent stage boundaries (denoted by their first three letters) associated with mass extinctions.

	Extinction				Origination			
Stage boundary	Percentage of decrease	P _{SOR}	P _{SOV}	P _{DPOC}	Percentage of increase	P _{SOR}	P _{SOV}	P _{DPOC}
Kat/Hir	48	+0.2	-3.9	1.3	16	-3.8	-25.2	3.4
Hir/Lla	24	-0.2	-0.9	0.8	618	+1.9	+7.1	1.6
Loc/Pra	41	-0.5	-1.8	0.8	31	-6.7	-7.9	2.2
Pra/Ems	41	-0.4	-2.4	0.9	21	-1.7	-15.8	2.7
Ems/Eif	70	+0.9	-10.4	1.8	22	-5.8	-22.0	3.1
Eif/Giv	33	+1.4	-4.8	1.2	0	_	_	-
Giv/Fra	13	+0.2	-1.3	0.8	0	_	_	-
Fra/Fam	14	+0.7	-2.2	0.9	200	-3.1	+8.7	2.7
Fam/Tou	50	+3.7	-5.6	1.3	20	± 0.0	-23.1	3.3
Kas/Gzh	58	+3.9	-10.5	2.0	17	-16.6	+2.0	4.8

Ordovician: Kat = Katian, Hir = Hirnantian; Silurian: Lla = Llandovery; Devonian: Loc = Lochkovian, Pra = Pragian, Ems = Emsian, Eif = Eifelian, Giv = Givetian, Fra = Frasnian, Fam = Famennian; Carboniferous: Tou = Tournasian, Kas = Kasimovian, Gzh = Gzhelian.

Values in regular font indicate changes due to extinction, whereas values in italics show changes due to origination. *P*_{SOV} quantifies the percentage difference in SOV between the first and second stage, *P*_{SOR} is the percentage difference in SOR, and *P*_{DPOC} is the percentage of change in the position of the centroid. See Table S11 for the raw values.

reduction of morphospace occupation in Eurypterina and Stylonurina. During the Devonian, Stylonurina and Eurypterina independently underwent an ecological transition from consisting of predominantly marine taxa to comprising purely freshwater species (O'Connell 1916; Lamsdell and Braddy 2010), while retaining their cosmopolitan distribution. This resulted in a decrease in diversity and an apparent stagnation in their evolution. It is possible that, due to their widespread distribution but limited founding pool of closely related species, each clade suffered a reduced morphological variability akin to the loss of species genetic variability due to small founder populations (Mayr 1963). Rapid recovery has been shown to somewhat mitigate such a loss in variability at the level of both populations (Nei et al. 1975) and clades, whereby the rate at which a group invades available ecological space after mass extinctions can greatly influence its chances of long-term recovery (Sahney et al. 2010b). Much of the late Paleozoic, however, is characterized by sluggish macroevolution, in which origination rates are depressed following the formation of glaciers in the Mississippian, resulting in species having broad ecological niches, widespread geographic distributions, and large and relatively stable populations (Stanley and Powell 2003; Powell 2005). Although some lineages underwent rapid recovery in the early Carboniferous (Smithson et al. 2016), prior to the onset of the Mississippian glaciation, eurypterids showed only a muted recovery. This may have been, at least in part, due to the fact that while post-Devonian eurypterids comprised two disparate clades (having diverged over 110 million years prior), each with distinct feeding ecologies, both were limited to freshwater environments (Lamsdell and Braddy 2010). Interestingly, xiphosurids are also shown to invade freshwater environments at this time (Lamsdell 2016), while the majority of the earliest Carboniferous vertebrate faunas are also known from nonmarine localities (Sallan and Coates 2010; Sallan and Galimberti 2015), and nonmarine environments may have acted as refugia for lineages impacted by the biotic crisis that has been suggested to be primarily limited to the pelagic realm (Racki 1999). For eurypterids, the

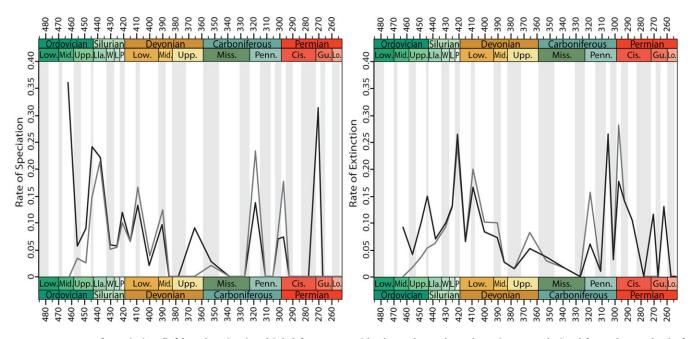


Figure 6. Rates of speciation (left) and extinction (right) for Eurypterida throughout the Paleozoic. Rates derived from the method of Foote (2000a,b) are shown in gray, those derived from the method of Hulbert (1993) in black.

limitation to nonmarine environments appears to have resulted in a reduction in the fundamental niche (the abiotic conditions in which an organism can survive: Pearman et al. 2008), with niche conservatism resulting in an inability to reoccupy the vacated niche space (Wiens 2004; Wiens et al. 2010) meaning that, unlike other groups, they would have been unable to successfully colonize vacant marine ecospace. The onset of sluggish macroevolution during the Mississippian, therefore, resulted in an apparently permanent loss in variability within the group. This resulted in a high degree of morphological stability (bradytely) with taxa such as Adelophthalmus and Cyrtoctenus remaining virtually unchanged for over 100 million years, a period of stasis equal to that exhibited by modern living fossils (stabilomorphs: Kin and Błażejowski 2014). It may be that extant stabilomorphs have similar evolutionary histories to those of the eurypterid clades studied here: hagfish and lampreys may exhibit far greater disparity in the Paleozoic than is evident among extant species (McCoy et al. 2016), whereas horseshoe crabs certainly exhibit greater morphological and ecological disparity during the Paleozoic and Mesozoic than they do today (Lamsdell 2016). This also has ramifications for modern conservation efforts because saving only a few species of a given clade is unlikely to result in long-term lineage survival or a return to its full breadth of previous ecological roles, lending support for the incorporation of phylogenetic diversity metrics into nature conservation studies (Winter et al. 2013).

The drop in diversity during the late Devonian is one of the five largest declines in the Phanerozoic (Raup and Sepkoski 1982; Sepkoski 1986); however, extinction rate actually decreased throughout the late Devonian (Orzechowski et al. 2015), and both

speciation and extinction rates decreased throughout the Devonian for Eurypterida (Fig. 6). Of the major late Devonian posited extinction events (House 1985, 2002), only the Hangenberg Event appears to be a real, abiotically driven mass extinction (Friedman and Sallan 2012), while the Kellwasser Event has been suggested to be a stratigraphic artifact of last occurrences caused by changes of water depth (Holland and Patzkowsky 2015), which characterize these horizons (McGhee 1996; Hallam and Wignall 1997). Instead, the long-term diversity decline appears to be due to decreased origination rather than elevated extinction (Bambach et al. 2004). It has been suggested that the drop in speciation rates was due to a decrease in endemism (May 1995; Stigall 2012), with a shutdown in vicariant speciation attributable to widespread interbasinal species invasions mediated by a global increase in sea level (Rode and Lieberman 2004; Abe and Lieberman 2009; Stigall 2010). Eurypterids show a high degree of endemism (Tetlie 2007b), possibly tied to their need to return to sheltered regions within very specific environmental tolerances to molt and possibly mate (Braddy 2001; Vrazo and Braddy 2011); similar population segregation is seen in modern horseshoe crabs, which show a strong correlation between genetic and geographic distance (King et al. 2005). Eurypterids show a marked drop in speciation from the Emsian, undergoing a recovery in origination rates during the Famennian. Other invertebrate groups in which a drop in speciation rates during the Devonian has been recorded-trilobites (Abe and Lieberman 2009), phyllocarids (Rode and Lieberman 2005), and brachiopods and bivalves (Stigall 2010)-are also highly endemic prior to the onset of the biotic crisis. However, the timing for the shutdown in speciation is different for each of these groups; speciation stops for some trilobite groups from the Emsian onward (Abe and Lieberman 2009) and phyllocarids exhibit low levels of speciation from the Eifelian with a slight recovery in the Frasnian (Rode and Lieberman 2005), whereas the speciation rates of the studied bivalve and brachiopod clades only tail off during the Frasnian and Famennian (Stigall 2010). The differential onset of depressed rates of speciation is likely due to variations in ecology resulting in differences in population structure (Fordham et al. 2013) and ability to track environmental changes (Saupe et al. 2014) that, when combined with regional geographic differences, mean different groups have different ecological thresholds beyond which speciation shuts down.

The origins of the Devonian biotic crisis may, therefore, be traced to the beginning of the period. Sea-level varied consistently throughout the Silurian and began to stabilize during the onset of the Devonian, remaining generally high throughout the period (Haq and Schutter 2008). The Silurian is characterized by multiple pulses of ocean anoxia and heavy metal excursions (Vandenbrouke et al. 2015), resulting in an unstable environment with a high degree of biotic turnover. This boom and bust macroevolution can maintain high standing diversity while the system is maintained; however, a reduction in rates can impact groups differently depending on whether they were in a boom or bust phase. Eurypterids appear to have ended the Silurian on a bust, experiencing marked extinction during the Pridoli followed by another in the Pragian (Fig. 6). Surviving clades were subsequently winnowed by background extinction throughout the remainder of the Devonian, with only a handful maintaining the rates of speciation required to persist, resulting in the onset of the shift in occupied morphospace as of the Middle Devonian. A similar shift in morphospace over the same timeframe is seen in crinoids (Foote 1994), which also begin their recovery during the Famennian (Sallan et al. 2011).

The Devonian biotic crisis had a marked impact on multiple taxonomic groups, interrupting or altering evolutionary trajectories and resulting in noticeable shifts in evolutionary regime. Vertebrates, for example, experienced body-size reduction in response to the Hangenberg Event (Sallan and Galimberti 2015). Post-Devonian eurypterids had a more global distribution than their Silurian ancestors (Tetlie 2007b), and trilobites exhibit a similar biogeographic pattern (Fortey and Owens 1997). The trends observed among the studied arthropod groups are mirrored in other metazoan clades (McGhee 1996); the biotic response to the Devonian biodiversity crisis appears to be characterized by global homogenization, with many losers and few winners. If the primary causal factors for this trend are a breakdown in endemism mediated by a rise in sea level, this has implications for marine conservation efforts given projected sea-level rise over the next 100 years (Meehl et al. 2005; Rahmstorf 2007; Vermeer and Rahmstorf 2009).

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DATA ARCHIVING

The doi for the study data is 10.5061/dryad.24767 on DRYAD and 10.7934/P2384 on Morphobank.

LITERATURE CITED

- Abe, F. R., and B. S. Lieberman. 2009. The nature of evolving radiations: a case study involving Devonian trilobites. Evol. Biol. 36:225–234.
- Alroy, J. 2010a. Fair sampling of taxonomic richness and unbiased estimation of origination and extinction rates. Pp. 55–80 *in* J. Alroy and G. Hunt, eds. Quantitative methods in paleobiology. Paleontological Society Papers 16. Paleontological Society and Geological Society of America. Boulder, Colorado.
- ——. 2010b. The shifting balance of diversity among major marine animal groups. Science 329:1191–1194.
- 2010c. Geographical, environmental and intrinsic biotic controls on Phanerozoic marine diversification. Palaeontology 53:1211–1235.
- 2014. Accurate and precise estimates of origination and extinction rates. Paleobiology 40:374–397.
- Anderson, R. P., V. E. McCoy, M. E. McNamara, and D. E. G. Briggs. 2014. What big eyes you have: the ecological role of giant pterygotid eurypterids. Biol. Lett. 10:1–4.
- Bambach, R. K., A. J. Knoll, and S. C. Wang. 2004. Origination, extinction, and mass depletions of marine diversity. Paleobiology 30:522– 542.
- Bapst, D. W. 2014. Assessing the effect of time-scaling methods on phylogenybased analyses in the fossil record. Paleobiology 40:331–351.
- Bapst, D. W., P. C. Bullock, M. J. Melchin, H. D. Sheets, and C. E. Mitchell. 2012. Graptoloid diversity and disparity became decoupled during the Ordovician mass extinction. Proc. Natl. Acad. Sci. USA 109:3428– 3433.
- Barnosky, A. D., N. Matzke, S. Tomiya, G. O. U. Wogan, B. Swartz, T. B. Quental, C. Marshall, J. L. McGuire, E. L. Lindsey, K. C. Maguire, et al. 2011. Has the Earth's sixth mass extinction already arrived? Nature 471:51–57.
- Bell, M. A., and G. T. Lloyd. 2015. strap: an *R* package for plotting phylogenies against stratigraphy and assessing their stratigraphic congruence. Palaeontology 58:379–389.
- Bond, D. P. G., and P. B. Wignall. 2008. The role of sea-level change and marine anoxia in the Frasnian-Famennian (Late Devonian) mass extinction. Palaeogeogr. Palaeoclimatol. Palaeoecol. 263:107–118.
- Boyer, D. L., Haddad, E. E., and Seeger, E. E. 2014. The last gasp: trace fossils track deoxygenation leading into the Frasnian-Famennian extinction event. Palaios 29:646–651.
- Braddy, S. J. 2000. Eurypterids from the Early Devonian of the Midland Valley of Scotland. Scot. J. Geol. 36:115–122.

— 2001. Eurypterid palaeoecology: palaeobiological, ichnological and comparative evidence for a "mass–moult–mate" hypothesis. Palaeogeogr. Palaeoclimatol. Palaeoecol. 172:115–132.

- Braddy, S. J., R. J. Aldridge, and J. N. Theron. 1995. A new eurypterid from the Late Ordovician Table Mountain Group, South Africa. Palaeontology 38:563–581.
- Brusatte, S. L., M. J. Benton, M. Ruta, and G. T. Lloyd. 2008. Superiority, competition, and opportunism in the evolutionary radiation of dinosurs. Science 321:1485–1488.
- Brusatte, S. L., S. Montanari, H-Y. Yi, and M. A. Norell. 2011. Phylogenetic corrections for morphological disparity analysis: a new methodology and case studies. Paleobiology 37:1–22.
- Budil, P., S.Manda, and O. E. Tetlie. 2014. Silurian carcinosomatid eurypterids from the Prague Basic (Czech Republic). Bull. Geosci. 89:257–267.
- Butler, R. J., P. M. Barrett, S. Nowbath, and P. Upchurch. 2009. Estimating the effects of sampling biases on pterosaur diversity patterns: implications for hypotheses of bird/pterosaur competitive replacement. Paleobiology 35:432–446.
- Cailliez, F. 1983. The analytical solution to the additive constant problem. Psychometrika 48:305–308.
- Carmichael, S. K., J. A. Waters, C. J. Batchelor, D. M. Coleman, T. J. Suttner, E. Kido, L. M. Moore, and L. Chadimová. 2016. Climate instability and tipping points in the Late Devonian: detection on the Hangenberg Event in an open oceanic island arc in the Central Asian Orogenic Belt. Gondwana Res. 32:213–231.
- Caster, K. E., and E. N. Kjellesvig-Waering. 1964. Upper Ordovician eurypterids of Ohio. Palaeontogr. Am. 4:1–358.
- Cavin, L., and P. L. Forey. 2007. Using ghost lineages to identify diversification events in the fossil record. Biol. Lett. 3:201–204.
- Chen, D., H. Quing, and R. Li. 2005. The late Devonian Frasnian–Famennian (F/F) biotic crisis: insights from δ;¹³C_{carb}, δ;¹³C_{org} and ⁸⁷Sr/⁸⁶Sr isotopic systematics. Earth and Planet. Sci. Lett. 235:151–166.
- Cohen, J. 1988. Statistical power analysis for the behavioural sciences. 2nd ed. Lawrence Erlbaum Associates, Hillsdale, NJ.
- Congreve, C. R. 2013. Cladal turnover: the end-Ordovician as a large-scale analogue of species turnover. Palaeontology 56:1285–1296.
- Congreve, C. R., and J. C. Lamsdell. 2016. Implied weighting and its utility in palaeontological datasets: a study using modelled phylogenetic matrices. Palaeontology 59:447–462.
- Cope, E. D. 1886. An interesting connecting genus of Chordata. Am. Nat. 1885:1027–1031.
- Dommergues, J. L., B. Laurin, and C. Meister. 1996. Evolution of ammonoid morphospace during the Early Jurassic radiation. Paleobiology 22:219– 240.
- Dunlop, J. A., D. Penney, and D. Jekel. 2015. A summary list of fossil spiders and their relatives. World Spider Catalog. Natural History Museum, Bern. Available at http://wsc.nmbe.ch.
- Foote, M. 1991. Morphological and taxonomic diversity in a clade's history: the blastoid record and stochastic simulations. Contrib. Mus. Paleontol. Univ. Mich. 28:101–140.
 - —. 1994. Morphological disparity in Ordovician-Devonian crinoids and the early saturation of morphological space. Paleobiology 20:320–344.
 - . 1999. Morphological diversity in the evolutionary radiations of Paleozoic and post-Paleozoic crinoids. Paleobiology 25(supplement):1–115.
- 2000a. Origination and extinction components of taxonomic diversity: general problems. Paleobiology 26:74–102.
- 2000b. Origination and extinction components of taxonomic diversity: Paleozoic and post-Paleozoic dynamics. Paleobiology 26:578–604.
- Fordham, D. A., C. Mellin, B. D. Russell, R. H. Akçakaya, C. J. A. Bradshaw, M. E. Aiello-Lammens, J. M. Caley, S. D. Connell, S. Mayfield, S. A. Shepherd, et al. 2013. Population dynamics can be more important than

physiological limits for determining range shifts under climate change. Glob. Chang. Biol. 19:3224–3237.

- Fortey, R. A., and R. M. Owens. 1997. Evolutionary history. Pp. 250–287 in R. L. Kaesler, ed. Treatise on invertebrate paleontology, part O, Arthropoda 1, Trilobita, Revised. Volume 1: introduction, order Agnostida, order Redlichiida. Geological Society of America and University of Kansas. Boulder, Colorado, and Lawrence, KS.
- Foth, C., S. L. Brusatte, and R. J. Butler. 2012. Do different disparity proxies converge on a common signal? Insights from the cranial morphometrics and evolutionary history of Pterosauria (Diaspsida: Archosauria). J. Evol. Biol. 25:904–915.
- Friedman, M., and L. C. Sallan. 2012. Five hundred million years of extinction and recovery: a Phanerozoic survey of large-scale diversity patterns in fishes. Palaeontology 55:707–742.
- Garamszegi, L. 2006. Comparing effect sizes across variables: generalization without the need for Bonferroni correction. Behav. Ecol. 17:682– 687.
- George, A. D., N. Chow, and K. M. Trinajstic. 2014. Oxic facies and the Late Devonian mass extinction, Canning Basin, Australia. Geology 42:327– 330.
- Goloboff, P. A., J. A. Farris, and K. C. Nixon. 2008. TNT, a free program for phylogenetic analysis. Cladistics 24:774–786.
- Gower, J. C. 1971. A general coefficient of similarity and some of its properties. Biometrics 27:857–871.
- Hallam, A., and P. B. Wignall. 1997. Mass extinctions and their aftermath. Oxford Univ. Press, New York.
- Haq, B. U., and S. R. Schutter. 2008. A chronology of Paleozoic sea-level changes. Science 322:64–68.
- Hetherington, A. J., E. Sherratt, M. Ruta, M. Wilkinson, B. Deline, and P. C. J. Donoghue. 2015. Do cladistic and morphometric data capture common patterns of morphological disparity? Palaeontology 58:393–399.
- Holland, S. M., and M. E. Patzkowsky. 2015. The stratigraphy of mass extinction. Palaeontology 58:903–924.
- Hopkins, M. J., and A. B. Smith. 2015. Dynamic evolutionary change in post-Paleozoic echinoids and the importance of scale when interpreting changes in rates of evolution. Proc. Natl. Acad. Sci. USA 112:3758– 3763.
- House, M. R. 1985. Correlation of mid-Palaeozoic ammonoid evolutionary events with global sedimentary perturbations. Nature 313:17–22.
- 2002. Strength, timing, setting and cause of mid-Palaeozoic extinctions. Palaeogeogr. Palaeoclimatol. Palaeoecol. 181:5–25.
- Hughes, M., S. Gerber, and M. A. Wills. 2013. Clades reach highest morphological disparity early in their evolution. Proc. Natl. Acad. Sci. USA 110:13875–13879.
- Hulbert, R. C. 1993. Taxonomic evolution in North American Neogene horses (subfamily Equinae): the rise and fall of an adaptive radiation. Paleobiology 19:216–234.
- Hull, P. M., S. A. F. Darroch, and D. H. Erwin. 2015. Rarity in mass extinctions and the future of ecosystems. Nature 528:346–351.
- Jablonski, D. 1986. Causes and consequences of mass extinctions: a comparative approach. Pp. 313–329 in D. K. Elliott, ed. Dynamics of extinction. Wiley, New York.
- 2001. Lessons from the past: evolutionary impacts of mass extinctions. Proc. Natl. Acad. Sci. USA 98:5393–5398.
- 2002. Survival without recovery after mass extinctions. Proc. Natl. Acad. Sci. USA 99:8139–8144.
- _____. 2005. Mass extinctions and macroevolution. Paleobiology 31:192– 210.
- Kaiser, S. I., M. Aretz, and R. T. Becker. 2016. The global Hangenberg Crisis (Devonian–Carboniferous transition): review of a first-order mass extinction. Geol. Soc. London Spec. Publ. 423:387–437.

- Kin, A., and B. Błażejowski. 2014. The horseshoe crab of the genus *Limulus*: living fossil or stabilomorph? PLoS ONE 9:1–11.
- King, T. L., M. S. Eackles, A. P. Spidle, and H. J. Brockmann. 2005. Regional differentiation nad sex-biased dispersal among populations of the horseshoe crab *Limulus polyphemus*. Trans. Am. Fish. Soc. 134:441–465.
- Kjellesvig-Waering, E. N. 1950. A new Silurian *Hughmilleria* from West Virginia. J. Paleontol. 24:226–228.
- ———. 1961. The Silurian Eurypterida of the Welsh Borderland. J. Paleontol. 35:789–835.
- Korn, D., M. J. Hopkins, and S. A. Walton. 2013. Extinction space—a method for the quantification and classification in changes in morphospace across extinction boundaries. Evolution 67:2795–2810.
- Lamsdell, J. C. 2011. The eurypterid *Stoermeropterus conicus* from the lower Silurian of the Pentland Hills, Scotland. Monogr. Palaeontogr. Soc. 165:1–84.
- 2013. Redescription of *Drepanopterus pentlandicus* Laurie, 1892, the earliest known mycteropoid (Chelicerata: Eurypterida) from the early Silurian (Llandovery) of the Pentland Hills, Scotland. Earth Environ. Sci. Trans. R. Soc. Edinb. 103:77–103.
- 2016. Horseshoe crab phylogeny and independent colonisations of freshwater: ecological invasion as a driver for morphological innovation. Palaeontology 59:181–194.
- Lamsdell, J. C., and S. J. Braddy. 2010. Cope's rule and Romer's theory: patterns of diversity and gigantism in eurypterids and Palaeozoic vertebrates. Biol. Lett. 6:265–269.
- Lamsdell, J. C., and P. A. Selden. 2013. Babes in the wood—a unique window into sea scorpion ontogeny. BMC Evol. Biol. 13:1–46.
- Lamsdell, J. C., S. J. Braddy, and O. E. Tetlie. 2009. Redescription of *Drepanopterus abonensis* (Chelicerata: Eurypterida: Stylonurina) from the Late Devonian of Portishead, UK. Palaeontology 52:1113–1139.
- Lamsdell, J. C., İ. Hoşgör, and P. A. Selden. 2013a. A new Ordovician eurypterid (Arthropoda: Chelicerata) from southeast Turkey: evidence for a cryptic Ordovician record of Eurypterida. Gondwana Res. 23:354–366.
- Lamsdell, J. C., L. Simonetto, and P. A. Selden. 2013b. First eurypterid from Italy: a new species of *Adelophthalmus* (Chelicerata: Eurypterida) from the Upper Carboniferous of the Carnic Alps (Friuli, NE Italy). Riv. Ital. Paleontol. S. 119:147–151.
- Lamsdell, J. C., D. E. G. Briggs, H. P. Liu, B. J. Witzke, and R. M. McKay. 2015. The oldest described eurypterid: a giant Middle Ordovician (Darriwilian) megalograptid from the Winneshiek Lagerstätte of Iowa. BMC Evol. Biol. 15:1–31.
- Lane, A., C. M. Janes, and J. J. Sepkoski Jr. 2005. Estimating paleodiversities: a test of the taxic and phylogenetic methods. Paleobiology 31:21–34.
- Leutze, W. P. 1961. Arthropods from the Syracuse Formation, Silurian of New York. J. Paleontol. 35:49–64.
- Levinton, J. 1988. Genetics, paleontology, and macroevolution. Cambridge Univ. Press, Cambridge, U.K.
- Liggs, U., and M. Maeler. 2003. Scatterplot3d an R package for visualizing multivariate data. J. Stat. Softw. 8:1–20.
- Lloyd, G. T. 2016. Estimating morphological diversity and tempo with discrete character-taxon matrices: implementation, challenges, progress, and future directions. Biol. J. Linn. Soc. 118:131–151.
- Lofgren, A. S., R. E. Plotnick, and P. J. Wagner. 2003. Morphological diversity of Carboniferous arthropods and insights on disparity patterns through the Phanerozoic. Paleobiology 29:349–368.
- Long, J. A., R. R. Large, M. D. Y. Lee, M. J. Benton, L. V. Danyushevsky, L. M. Chiappe, J. A. Halpin, D. Cantrill, and B. Lottermoser. 2016. Severe selenium depletion in the Phanerozoic oceans as a factor in three global mass extinction events. Gondwana Res. 36:209–218.
- Ma, X., Y. Gong, D. Chen, G. Racki, X. Chen, and W. Liao. 2016. The late Devonian Frasnian-Famennian Event in South China—patterns and causes

of extinctions, sea level changes, and isotope variations. Palaeogeogr. Palaeoclimatol. Palaeoecol. 448:224–244.

- Maddison, W. P., and D. R. Maddison. 2015. Mesquite: a modular system for evolutionary analysis. Version 3.04. Available at http:// mesquiteproject.org
- Maurer, B. A. 1989. Diversity-dependent species dynamics: incorporating the effects of population-level processes on species dynamics. Paleobiology 15:133–146.
- May, A. 1995. Relationship among sea-level fluctuation, biogeography and bioevents of the Devonian: an attempt to approach a powerful, but simple model for complex long-range control of biotic crises. GeoLines 3:38– 49.
- Mayr, E. 1963. Animal species and evolution. Harvard Univ. Press, Cambridge, MA.
- McCoy, V. E., J. C. Lamsdell, M. Poschmann, R. P. Anderson, and D. E. G. Briggs. 2015. All the better to see you with: eyes and claws reveal the evolution of divergent ecological roles in giant pterygotid eurypterids. Biol. Lett. 11:1–4.
- McCoy, V. E., E. E. Saupe, J. C. Lamsdell, L. G. Tarhan, S. McMahon, S. Lidgard, P. Mayer, C. D. Whalen, C. Soriano, L. Finney, et al. 2016. The "Tully monster" is a vertebrate. Nature 532:496–499.
- McGhee, G. R. 1996. The Late Devonian mass extinction. Columbia Univ. Press, New York.
- McGhee, G. R., M. E. Clapham, P. M. Sheehan, D. J. Bottjer, and M. L. Droser. 2013. A new ecological-sensitivity ranking of major Phanerozoic biodiversity crises. Palaeogeogr. Palaeoclimatol. Palaeoecol. 370:260– 270.
- McKinney, M. L., and L. K. Lockwood. 1999. Biotic homogenization: a few winners replacing many losers in the next mass extinction. Trends Ecol. Evol. 14:450–453.
- Meehl, G. A., W. M. Washington, W. D. Collins, J. M. Arblaster, A. Hu, L. E. Buja, W. G. Strand, and H. Teng. 2005. How much more global warming and sea level rise? Science 307:1769–1772.
- Moran, M. D. 2003. Arguments for rejecting the sequential Bonferroni in ecological studies. Oikos 100:403–405.
- Myrow, P. M., J. Ramezani, A. E. Hanson, S. A. Bowring, G. Racki, and M. Rakociński. 2014. High-precision U-Pb age and duration of the latest Devonian (Famennian) Hangenberg event, and its implications. Terra Nova 26:222–229.
- Nakagawa, S. 2004. A farewell to Bonferroni: the problems of low statistical power and publication bias. Behav. Ecol. 15:1044–1045.
- Nee, S. 2004. Extinct meets extant: simple models in paleontology and molecular phylogenetics. Paleobiology 30:172–179.
- 2006. Birth-death models in macroevolution. Ann. Rev. Ecol. Evol. Syst. 37:1–17.
- Nei, M., T. Msruyama, and R. Chakraborty. 1975. The bottleneck effect and genetic variability in populations. Evolution 29:1–10.
- Newell, N. D. 1959. Adequac of the fossil record. J. Paleontol. 26:371-385.
- Norell, M. A. 1992. Taxic origin and temporal diversity: the effect of phylogeny. Pp. 89–118 in M. Novacek, and Q. Wheeler, eds. Extinction and phylogeny. Columbia Univ. Press, New York.
- . 1993. Tree-based approaches to understanding history: comments on ranks, rules, and the quality of the fossil record. Am. J. Sci. 293:407– 417.
- O'Connell, M. 1916. The habitat of the Eurypterida. Bull. Buffalo Soc.Nat. Sci. 11, 1–277.
- O'Leary, M. A., and S. G. Kaufman. 2012. MorphoBank 3.0: web application for morphological phylogenetics and taxonomy. Available at http://www.morphobank.org.
- Orzechowski, E. A., R. Lockwood, J. E. K. Byrnes, S. C. Anderson, S. Finnegan, Z. V. Finkel, P. G. Harnik, D. R. Lindberg, L. H. Liow, H. K.

Lotze, et al. 2015. Marine extinction risk shaped by trait-environment interactions over 500 million years. Glob. Change Biol. 21:3595–3607.

- Oyston, J. W., M. Hughes, S. Gerber, and M. A. Wills. 2016. Why should we investigate the morphological disparity of plant clades? Ann. Bot. 117:859–879.
- Patterson, C., and A. B. Smith. 1989. Periodicity in extinction: the role of systematics. Ecology 70:802–811.
- Pearman, P. B., A. Guisan, O. Broennimann, and C. F. Randin. 2008. Niche dynamics in space and time. Trends Ecol. Evol. 23:149–158.
- Plotnick, R. E., F. A. Smith, and S. K. Lyons. 2016. The fossil record of the sixth extinction. Ecol. Lett. 19:546–553.
- Powell, M. P. 2005. Climatic basis for suggish macroevolution during the late Paleozoic ice age. Geology 33:381–384.
- Racki, G. 1999. The Frasnian–Famennian biotic crisis: how many (if any) bolide impacts? Geol. Runsdch. 87:617–632.
- Rahmstorf, S. 2007. A semi-empirical approach to projecting future sea-level rise. Science 315:368–370.
- Rakociński, M., A. Pisarzowska, K. Janiszewska, and P. Szrek. 2016. Depositional conditions during the Lower Kellwasser Event (Late Frasnian) in the deep-sheld Łysogóry Basin of the Holy Cross Mountains Poland. Lethaia 49:571–590.
- Raup, D. M. 1972. Taxonomic diversity during the Phanerozoic. Science 177:1065–1071.

- Raup, D. M., and J. J. Sepkoski. 1982. Mass extinctions in the marine fossil record. Science 215:1501–1502.
- Retallack, G. J. 2011. Exceptional fossil preservation during CO₂ greenhouse crises? Palaeogeogr. Palaeoclimatol. Palaeoecol. 307:59–74.
- Rode, A. L., and B. S. Lieberman. 2004. Using GIS to unlock the interactions between biogeography, environment, and evolution in Middle and Late Devonian brachiopods and bivalves. Palaeogeogr. Palaeoclimatol. Palaeoecol. 211:345–359.

——. 2005. Integrating evolution and biogeography: a case study involving Devonian crustaceans. J. Paleontol. 79:267–276.

- Rolfe, W. D. I. 1963. Morphology of the telson of *Ceratiocaris cornwallisensis* (Crustacea: Phyllocarida) from Czechoslovakia. J. Paleontol. 37:486– 488.
- Ruta, M., K. D. Angielczyk, J. Fröbisch, and M. J. Benton. 2013. Decoupling of morphological disparity and taxic diversity during the adaptive radiation of anomodont therapsids. Proc. R. Soc. B 280:1–9.
- Sahney, S., M. J. Benton, and H. J. Falcon-Lang. 2010a. Rainforest collapse triggered Carboniferous tetrapod diversification. Geology 38:1079– 1082.
- Sahney, S., M. J. Benton, and P. A. Ferry. 2010b. Links between global taxonomic diversity, ecological diversity and the expansion of vertebrates on land. Biol. Lett. 6:544–547.
- Sallan, L. C., and M. I. Coates. 2010. End-Devonian extinction and a bottleneck in the early evolution of modern jawed vertebrates. Proc. Natl. Acad. Sci. USA 107:10131–10135.
- Sallan, L. C., and A. K. Galimberti. 2015. Body-size reduction in vertebrates following the end-Devonian mass extinction. Science 350:812–815.
- Sallan, L. C., T. W. Kammer, W. I. Ausich, and L. A. Cook. 2011. Persistent predator–prey dynamics revealed by mass extinction. Proc. Natl. Acad. Sci. USA 108:8335–8338.
- Sanderson, M. J., and M. J. Donoghue. 1996. Reconstructing shifts in diversification rates on phylogenetic trees. Trends Ecol. Evol. 11:15–20.
- Saupe, E. E., J. R. Hendricks, R. W. Portell, H. J. Dowsett, A. Haywood, S. J. Hunter, and B. S. Lieberman. 2014. Macroevolutionary consequences

of profound climate change on niche evolution in marine molluscs over the past three million years. Proc. R. Soc. B 281:1–9.

- Selden, P. A. 1986. A new identity for the Silurian arthropod Necrogammarus. Palaeontology 29:629–631.
- Sepkoski, J. J. 1986. Phanerozoic overview of mass extinction. Pp. 277–285 in D. M. Raup, and J. J. Sepkoski, eds. Patterns and processes in the history of life. Springer-Verlag, Berlin.
- Sheehan, P. M. 2001. The Late Ordovician mass extinction. Annu. Rev. Earth Planet. Sci. 29:331–364.
- Signor, P. W., and J. H. Lipps. 1982. Sampling bias, gradual extinction patterns and catastrophes in the fossil record. Geol. Soc. Spec. Pap. 190:291–296.
- Simpson, G. G. 1960. The history of life. Pp. 117–180 in S. Tax, ed. Evolution after Darwin. Volume 1: the evolution of life. Univ. of Chicago Press, Chicago, IL. 629 p.
- Slavík, L., J. I. Valenzuela-Ríos, J. Hladil, L. Chadimová, J.-C. Liao, A. Hušková, H. Calvo, and T. Hrstka. 2016. Warming or cooling in the Pragian? Sedimentary record and petrophysical logs across the Lochkovian-Pragian boundary in the Spanish Central Pyrenees. Palaeogeogr. Palaeoclimatol. Palaeoecol. 449:300–320.
- Smith, A. B. 1994. Systematics and the fossil record. Blackwell Scientific, Oxford, U.K.
- Smithson, T. R., K. R. Richards, and J. A. Clack. 2016. Lungfish diversity in Romer's Gap: reaction to the end-Devonian extinction. Palaeontology 59:29–44.
- Sokal, R. R., and F. J. Rohlf. 1995. Biometry. 3rd ed. W. H. Freeman and Company, New York.
- Stanley, S. M., and M. G. Powell. 2003. Depressed rates of origination and extinction during the late Paleozoic ice age: a new state for the global marine ecosystem. Geology 31:877–880.
- Stigall, A. L. 2010. Invasive species and biodiversity crises: testing the link in the Late Devonian. PLoS ONE 5:1–7.
- 2012. Speciation collapse and invasive species dynamics during the Late Devonian "Mass Extinction." GSA Today 22:4–9.
- Størmer, L. 1973. Arthropods from the lower Devonian (Lower Emsian) of Alken an der Mosel, Germany. Part 3: Eurypterida, Hughmilleriidae. Senck. Leth. 54:119–205.
- 1974. Arthropods from the Lower Devonian (Lower Emsian) of Alken an der Mosel, Germany. Part 4: Eurypterida, Drepanopteridae, and other groups. Senck. Leth. 54:359–451.
- Tetlie, O. E. 2006. Eurypterida (Chelicerata) from the Welsh Borderlands, England. Geol. Mag. 143:723–735.
- 2007b. Distribution and dispersal history of Eurypterida (Chelicerata). Palaeogeogr. Palaeoclimatol. Palaeoecol. 252:557–574.
- Tetlie, O. E., and P. Van Roy. 2006. A reappraisal of *Eurypterus dumonti* Stainier, 1917 and its position within the Adelophthalmidae Tollerton, 1989. Bull. Inst. R. Sci. Nat. Belg. Sci. Terr. 76:79–90.
- Tetlie, O. E., V. P. Tollerton Jr., and S. J. Ciurca Jr. 2007. *Eurypterus remipes* and *E. lacustris* (Chelicerata: Eurypterida) from the Silurian of North America. Bull. Peabody Mus. Nat. Hist. 48:139–152.
- Thorne, P. M., M. Ruta, and M. J. Benton. 2011. Resetting the evolution of marine reptiles at the Triassic-Jurassic boundary. Proc. Natl. Acad. Sci. USA 108:8339–8344.
- Van Valen, M. 1974. Multivariate statistics in natural history. J. Theor. Biol. 45:235–247.
- Vandenbroucke, T. R. A., P. Emsbo, A. Munnecke, N. Nuns, L. Duponchel, K. Lepot, M. Quijada, F. Paris, T. Servais, and W. Kiessling. 2015. Metalinduced malformations in early Palaeozoic plankton are harbingers of mass extinction. Nat. Commun. 6:1–7.

- Vermeer, M., and S. Rahmstorf. 2009. Global sea level linked to global temperature. Proc. Natl. Acad. Sci. USA 106:21527–21532.
- Vrazo, M. B., and S. J. Braddy. 2011. Testing the "mass-moult-mate" hypothesis of eurypterid palaeoecology. Palaeogeogr. Palaeoclimatol. Palaeoecol. 311:63–73.
- Wagner, P. J. 2000. The quality of the fossil record and the accuracy of phylogenetic inferences about sampling and diversity. Syst. Biol. 49:65– 86.
- Wang, S. C., A. E. Zimmerman, B. S. McVeigh, P. J. Everson, and H. Wong. 2012. Confidence interviews for the duration of mass extinction. Paleobiology 38:265–277.
- Wesley-Hunt, G. D. 2005. The morphological diversification of carnivores in North America. Paleobiology 31:35–55.
- Wiens, J. J. 2004. Speciation and ecology revisited: phylogenetic niche conservatism and the origin of species. Evolution 58:193–197.
- Wiens, J. J., D. D. Ackerly, A. P. Allen, B. L. Anacker, L. B. Buckley, H. V. Cornell, E. I. Damschen, T. J. Davies, J.-A. Grytnes, S. P. Harrison, et al. 2010. Niche conservatism as an emerging principle in ecology and conservation biology. Ecol. Lett. 13:1310–1324.

- Wills, M. A. 1998. Crustacean disparity through the Phanerozoic: comparing morphological and stratigraphic data. Biol. J. Linn. Soc. 65:455–500.
- Wills, M. A., D. E. G. Briggs, and R. A. Fortey. 1994. Disparity as an evolutionary index: a comparison of Cambrian and Recent arthropods. Paleobiology 20:93–130.
- Winter, M., V. Devictor, and O. Schweiger. 2013. Phylogenetic diversity and nature conservation: where are we? Trends Ecol. Evol. 28:199– 204.
- Young, M. T., S. L. Brusatte, M. Ruta, and M. B. De Andrade. 2010. The evolution of Metriorhynchoidea (Mesoeucrocodylia, Thalattosuchia): an integrated approach using geometric morphometrics, analysis of disparity, and biomechanics. Zool. J. Linn. Soc. 158:801–859.
- Zatoń, M., P. Filipiak, M., Rakociński, and W. Krawczyński 2014. Kowala Lagerstätte: late Devonian arthropods and non-biomineralized algae from Poland. Lethaia 47:352–364.

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Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

Table S1. Stage-level counts for species diversity, showing both taxic (observed) and phylogenetic (modeled) diversity.

Table S2. Summary of changes in the sum of range (SOR), sum of variance (SOV), and position of the centroid (DPOC) measured as percentages for random, lateral, and marginal variations in morphospace under 50% (regular font), 75% (italicized font), and 87.5% (bold font) change as reported from the simulations of Korn et al. (2013).

Table S3. PERMANOVA test results of Eurypterida (F(5,321) = 1.298, $\eta 2 = 0.016$, P = 0.0001) from 100,000 permutations with Bonferroni correction for statistical differences between taxa for each of the six period-level time bins based on PCO analyses.

Table S4. PERMANOVA test results of Eurypterida (F(14,494) = 1.092, $\eta 2 = 0.028$, P = 0.0003) from 100,000 permutations with Bonferroni correction for statistical differences between taxa for each of the 14 epoch-level time bins based on PCO analyses.

Table S5. PERMANOVA test results of Eurypterida (F(5,159) = 1.210, $\eta 2 = 0.028$, P = 0.0027) from 100,000 permutations with Bonferroni correction for statistical differences between taxa for each of the six period-level time bins based on PCO analyses, excluding inferred ancestral taxa.

Table S6. PERMANOVA test results of Eurypterida (F(14,257) = 0.976, $\eta 2 = 0.042$, P = 0.7527) from 100,000 permutations with Bonferroni correction for statistical differences between taxa for each of the 14 epoch-level time bins based on PCO analyses, excluding inferred ancestors.

Table S7. PERMANOVA test results of Eurypterida (F(14,494) = 1.092, $\eta 2 = 0.028$, P = 0.0003) from 100,000 permutations for statistical differences between taxa for each of the 14 epoch-level time bins based on PCO analyses.

Table S8. PERMANOVA test results of Eurypterida (F(5,159) = 1.210, $\eta 2 = 0.028$, P = 0.0027) from 100,000 permutations for statistical differences between taxa for each of the six period-level time bins based on PCO analyses, excluding inferred ancestral taxa.

Table S9. PERMANOVA test results of Eurypterida (F(14,257) = 0.976, $\eta 2 = 0.042$, P = 0.7527) from 100,000 permutations for statistical differences between taxa for each of the 14 Epoch-level time bins based on PCO analyses, excluding inferred ancestors.

Table S10. Quantification of changes in morphospace excluding inferred ancestors across stage boundaries (Korn et al. 2013).

Table S11. Quantification of changes in morphospace across stage boundaries (Korn et al. 2013), raw values.

Table S12. Quantification of changes in morphospace excluding inferred ancestors across stage boundaries (Korn et al. 2013), raw values.

Figure S1. Phylogenetic tree of the Eurypterida.

Figure S2. Disparity metrics for Eurypterida from the Euclidean distance matrix derived from the character matrix, excluding inferred ancestors.